

**TIER I AQUATIC VEGETATION
RECONNAISSANCE SURVEY PROTOCOL**

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Tier I Reconnaissance Survey Protocol

1.0 Strategy for Surveying Aquatic Vegetation

This procedure was developed to serve as a **qualitative** surveying mechanism for aquatic plants. This protocol will serve to meet the following objectives:

1. to provide a distribution map of the aquatic plant species within a waterbody
2. to document gross changes in the extent of a particular plant bed or the relative abundance of a species within a waterbody

This survey strategy may be augmented with the Tier II Aquatic Vegetation Sampling Protocol to gain more quantitative data if desired.

1.1 Introduction to Aquatic Plant Surveying

Surveys of aquatic vegetation are important to managers and researchers for habitat inventories, diagnosis of problem areas, detection of nuisance and/or exotic species, and in the development of aquatic vegetation management plans. Many levels of surveys may be completed from visual observation to highly quantitative, repeatable strategies. The major advantage of visual qualitative survey methods is the relatively small amount of time required to complete a survey. The standardization of a reconnaissance survey procedure will allow for more precise information to be collected. This increase in precision will occur as standardization allows many people to produce more similar results over time. The methods described below are designed to provide a standard rapid assessment of aquatic plant communities within a given waterbody. The following text, tables, figures and datasheets will strive to meet the goal of standardization. A quick and easy reconnaissance protocol may increase the number of repeatable future surveys leading to a greater understanding of aquatic plant distributions and changes within the state.

1.2 Interpreting Reconnaissance Surveys

Distribution maps generated from a Reconnaissance Survey may be converted to surface area information using standard map measuring techniques (i.e., compensating polar planimeter, digital software, or scaled grids). Measurement of surface area allows for rough quantification of the information collected during the survey. These data may be compared between species, seasons, and years to allow for development of conservation and management plans. However, it is important to remember that visual measures of abundance are highly subjective and data are more valuable if subjectivity can be reduced by better defining commonly used abundance descriptions (e.g., "rare" = <2% of the community). For this reason, in water bodies with low water clarity that prevents visual identification of plant species, rake throws are conducted to allow species identification.

1.3 Habitat Stratification

The types of areas/waterbodies commonly surveyed are divided into strata and subjected to discrete surveying efforts to increase efficiency, effectiveness, and knowledge of habitat influence on plant communities. Each stratum represents a major aquatic geomorphic feature in the State of Indiana (Table 1). A few other strata are not typically surveyed. The main navigation channel on the Ohio River and other deepwater areas within selected lakes or rivers (>6 m deep) are not surveyed because aquatic vegetation is unlikely to grow in these areas in the prevailing water quality conditions. In addition, the aquatic areas near dams and/or spillways are not surveyed because of safety considerations. Refer to Table 1 when categorizing the surveyed stratum.

Table 1. Aquatic Area Strata and Codes

Stratum Description	Stratum Code
Inland Lake	IL
Inland Reservoir	IR
Lake Michigan	LM
First Order Stream	FOS
Second Order Stream	SOS
Third Order Stream	TOS
Fourth Order Stream	FROS
Fifth Order Stream	FHOS
None	NA

* When “None” is selected, describe the habitat type in the comments section of the data sheet.

2.0 Equipment

2.1 Maps

A high-resolution bathymetric map is used as the base map when available. Potential map sources also include: printouts from digital sources, USGS 7.5 minute topographic maps, aerial photos, production of a map with a stadia rod and sighting compass, or a hand drawn sketch of the lake.

2.2 Field Equipment and Explanation

- A. Boat
- B. Safety Equipment (e.g., life jackets)
- C. Frodis (i.e., rake, anchor, or other sampling device w/ rope)
- D. Lake map
- E. Waterproof pens, pencils, or markers
- F. Plastic bags, cooler, and gel packs for collection of unknown plants
- G. Polarized sunglasses
- H. Secchi disk (optional)
- I. Range finder (optional)
- J. GPS unit (WAAS enabled)
- K. Depth detection device (e.g., sounding line, depth gun, sonar; optional)
- L. Aquaview (looking glass; optional)

A boat or canoe is needed to survey the lake. Any safety equipment that is required by law (U.S. Coast Guard or state law) should also be carried on board. A frodis is often needed to collect plants from deeper water areas and below canopies when visual inspection does not allow species determination. The lake map is marked with plant bed numbers and approximate boundaries to help distinguish plant beds of different species and/or groups of species within the lake. A unique datasheet will correspond to each plant bed number on the map for further explanation of that particular plant community. A Secchi disk will help determine the depth at which plants can be seen and will help define the littoral region. A range finder is helpful in determining distances from shore to more accurately map vegetation. WAAS-enabled GPS units should be used to determine locations of plant beds and perimeters of plant beds using tracks and/or waypoints. Coordinates may be uploaded to computers to map vegetation beds for permanent record and help in the determination of surface area of vegetation. Coordinates may also be plotted on scaled maps using map grids. A depth detection device may be used to determine the extent of the littoral region including shallow shoals offshore where aquatic vegetation may grow. Some sonar device models may also help determine the maximum depth of macrophyte growth. A looking glass may be used to more clearly see vegetation below the surface. Polarized sunglasses are a necessity as they greatly

improve one's ability to see below the surface and distinguish plants, thus reducing the number of rake tosses that are needed. Finally, plastic bags are on hand for the collection of unidentified species.

3. Preparation

3.1 Pre-survey Information

Prior to entering the field, information should be gathered on the lake being surveyed. Valuable information includes lake size, maximum depth, historical species lists (if available), and historical Secchi depth data. Sources for this information include the Indiana Department of Natural Resources' regional fisheries biologists, diagnostic study reports, websites (IDNR, IDEM, & IU), and other sources. The size and depth of the lake can help determine equipment needs and the amount of time needed to complete a survey of the lake. While survey time is often correlated to lake size, it is more closely related to the shoreline length and/or area of the littoral zone (i.e., large lakes with many coves have a greater shoreline length). **The acceptable sampling period extends from 15 June to 15 September. If resources are limited to a single reconnaissance survey, then the surveys should be conducted between 15 July and 31 August;** however, secondary surveys are recommended to catch temporal variations in plant communities. Also, depending on the intent of the survey, some *partial* lake surveys may be conducted.

3.2 Determination of Littoral Zone

The entire littoral zone of a lake should be briefly examined during a Reconnaissance Survey. Determination of the littoral zone is important for management and mapping of vegetative cover within a lake. The littoral zone is defined as the region of a lake from shore to a depth where vegetation disappears. In lakes with relatively shallow secchi depths the 1% light level may be approximated by multiplying the secchi depth by a factor of three. Most macrophyte species will not grow to the 1% light level, only algae and primitive plants. In extremely clear lakes macrophytes are generally restricted by hydrostatic pressure, rather than light, to a depth of 6 m (19.7 ft.) but some species may grow deeper. Eurasian watermilfoil has been found to grow to a depth of at least 9 m (30 ft.) while elodea has been found growing to a depth of 12 m. Isoetes (quillwort) has been found to grow to a depth of 15 m (~50 ft.) or more in clear lakes.

Secchi depth should be measured as follows:

- A. Anchor the boat to prevent drifting. Be careful not to disturb the sediments on the bottom when anchoring since this could cloud the water and interfere with the Secchi disk reading, especially in shallow lakes.
- B. Once you are at the deepest point of the lake, go to the shady side of the boat and if you are wearing sunglasses, remove them.
- C. Lower the Secchi disk (8-inch type) straight down into the water until the disk just disappears from sight. Mark the rope at the water level with a clothespin.
- D. Slowly raise the disk up until it reappears. Mark the rope at the water level with your fingers or with the other clothespin.
- E. To find the Secchi depth, grasp both clothespins in one hand and find the center of the loop of rope. Move one clothespin to that point and remove the other. This point is one-half the distance between the point of disappearance of the disk and the point where it re-appeared. Measure the distance from this point to the surface using a measuring tape.
- F. Record the Secchi depth on your data sheet to the nearest tenth of a foot.

The littoral zone of a lake, for purposes of a standard Reconnaissance Survey, is defined as the area from the shoreline to a depth equal to three times the known (or average) Secchi depths.

4. Surveying

4.1 Survey Coverage

Once the littoral zone of a lake has been determined, the survey can begin. The boat path should include a zig-zag pattern through the littoral region of the lake. Lakes that drop off quickly may only need one path along shore. In areas where the littoral region extends far from shore, several passes may need to be made in a zig-zag pattern. **These passes should never be farther apart than can be visually inspected.** For instance, if a bed of vegetation extends to the surface and it is visible from one side to the other, there is no need to make multiple passes through this area. However, in areas of dense canopies an effort is made to determine if any species are growing below the canopy. (This may include one to a few rake tosses.) Each unique plant bed requires a unique datasheet to be completed (see Appendix A). Any off-shore shoal areas that have a depth less than the maximum littoral zone depth are surveyed as well (an additional datasheet compiled for each). A photocopy of a bathymetric map for the lake with potential littoral areas outlined will be useful during field surveys to ensure that no areas are missed. A cover datasheet is completed for the waterbody as a whole and all individual plant bed datasheets are then attached to the waterbody coversheet.

Once approximately 50% of the shoreline areas are surveyed, a determination is made on the detail needed to survey within the remainder of the littoral region. It is important to sample areas that provide different habitat for plants (e.g., points, coves, shores with different features). Different shoreline attributes (e.g., face north, south, etc.) often contain different species (e.g., plants that sprout from fragments will often be more abundant on the windward side of a lake). In a lake with many species growing in relatively small beds, the littoral zone is examined more carefully than a lake with dense monoculture stands that cover large areas.

The time associated with a survey varies based upon factors noted above and the experience of the survey team. As much detail is collected as time allows. In general, most surveys completed using this protocol will take anywhere from three to eight hours to complete. The amount of time required is affected by the diversity of the plant beds and the amount of littoral region, more so than the lake size. Shoreline length also greatly affects the time needed to complete a Reconnaissance Survey. Generally, one to two miles of shoreline can be surveyed per hour. However, if the littoral region is narrow and/or diversity is low, a greater distance is surveyed per hour. Lakes less than 300 acres require approximately 2 hours per 100 acres. Lakes greater than 300 acres generally decrease in the time required per 100 acres. Lakes as large as 800 acres may be completed in one day. It is important to gather and review lake maps ahead of time since they provide the survey team with valuable information related to depth contours, shoreline length, and lake size; thus, allowing the team to devote an appropriate amount of time to the survey.

4.2 Vegetation Mapping & Data Recording

The survey technique utilizes a combination of intense visual examination and limited rake grabs to identify the abundance of aquatic species in individual plant beds. The individual plant bed survey area is defined as a contiguous, consistent (similar composition) community. This survey site/bed is then surveyed in its entirety. If the community composition changes dramatically while surveying what appears to be a contiguous bed, prepare individual datasheets for the different communities and note their approximate boundaries on the attached map.

A cover sheet is completed for each waterbody. Individual site data sheets for a given plant bed are then attached to the waterbody cover sheet. A new site data sheet is started for each plant bed in the waterbody. The survey sites/beds are numbered counter-clockwise around the waterbody beginning with "01", and remain the same from year to year (as much as possible). Information about each plant bed appears in its entirety on a single page. When there is not enough room remaining on a page to complete the listing for a bed,

a new page is started. **All numbered survey sites/beds have approximate boundaries sketched on a corresponding map and labeled with their unique number, such as “01”.**

The data sheet (Appendix A) is divided into four sections, **Site information**, **Site Coordinates**, **Species information**, and **Reminder information**. The **Reminder information** contains the data choices to be entered in the major data fields (boxes). All data fields on the data sheet are explained in detail in Appendix B.

The surveying operation is composed of multiple steps, beginning with recording **Species information**. Steps 1–4 are to collect **Species information**. Steps 5–8 are to complete **Site information and Site Coordinate information**.

Step 1. After a survey site/bed is reached, a site number, such as “01”, is recorded in the **Plant Bed ID** box of the **Site Information** section. This step signifies the beginning of the **Species Information** section. Travel in zig-zag pattern through the plant bed (See Figure 1). Record the species code (Appendix C) for all species in the **Species code** box and assign a visual abundance rating (in the **Abundance** box) for every submersed, rooted floating-leaved, non-rooted floating-leaved, emergent species, and alga observed. Assign abundance ratings based on the increments outlined in Table 2. These ratings essentially represent a percent cover measurement. If the canopy is dense or visibility is too poor for accurate visual identification of species, make sufficient rake throws to determine the occurrence of all species.

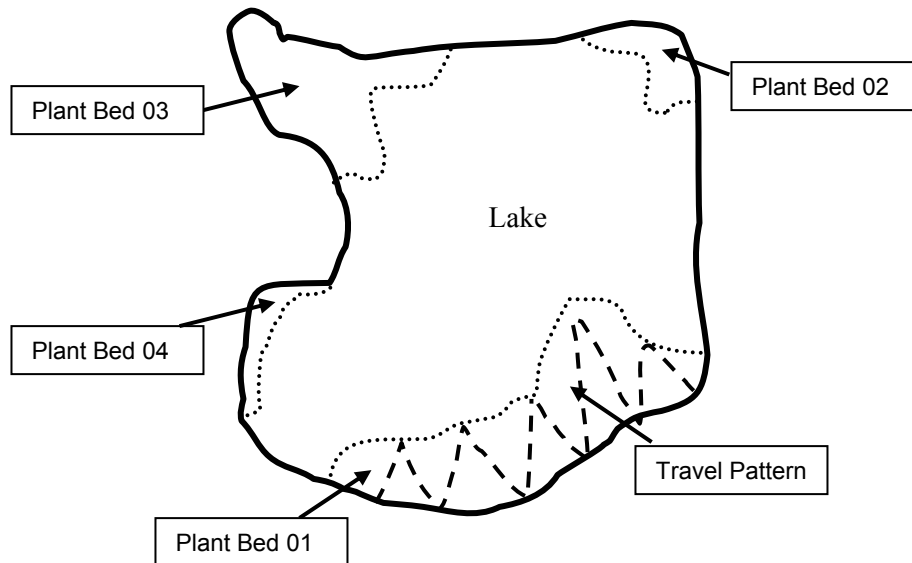


Figure 1: Depicts a potential travel path around a lake for purposes of a Reconnaissance Survey.

Table 2. Visual Abundance Ratings

Abundance (%)	Cover rating
> 61	4
21-60	3
2-20	2
< 2	1

Step 2. If the choice of species code fits the definition in Appendix B, assign a QE code of “0”. Otherwise, assign a code according to Table 3 in the **QE** box to flag each uncertain choice of species code.

Voucher specimens are collected for any species of which the identity is uncertain or unknown, or a species that is known not to be in the state herbarium.

Table 3. Plant identification quality evaluation codes

Identification certainty	QE code
Species code follows the definition in Appendix B	0
Genus certain, species suspected	1
Both genus and species suspected	2
unknown	3

Step 3. If a specimen is collected, a 1 is recorded in the **Voucher** box otherwise a “0” is recorded. If the specimen is sent to a taxonomist for identification, the 1 is later amended to a 2 to serve as a reminder that identification is pending. Comments about that site (unusual situations, species taken for identification or the presence of endangered, threatened, or rare (ETR) species) are written in the **Comments** section by the data recorder. If available the latitude and longitude location of any voucher specimens collected or the location of ETR species is also recorded in the comments sections. Voucher specimens should include multiple specimens of the same species (3-5 specimens with all available morphological characteristics, flowers, fruits, etc.)

Step 4. After reaching the perimeter of the plant bed, sketch the relative size and location of the individual bed on an attached lake map. Record the corresponding **Plant Bed ID** number on the map. It is also possible to assign a unique reference number/letter to denote the approximate location of a species of special interest on the map. Record this number/letter on both the map and the data sheet (**Ref. ID** box).

The area of the plant bed may be drawn onto the map with some accuracy if shoreline points of reference (e.g., points, docks, etc.) are used to determine your location on the lake. GPS units and rangefinders may also increase the accuracy of these sketches.

Step 5. After having surveyed the extent and composition of the plant bed, visually estimate by life form the percentage canopy cover of nonrooted floating-leaved, rooted floating-leaved, emergent, and submersed canopy species in the bed. Rate the percentage cover of the canopy species according to Table 4, and record the ratings in the appropriate **Canopy** box. (Note: Emergent, rooted floating-leaved, and nonrooted floating-leaved plants intercept sunlight at or above the water surface and may shade submersed plants growing in the water column, therefore, the percent canopy is important site information.) The rating should reflect the abundance of these life forms throughout the entire plant bed and serve to summarize the canopy cover and composition for the bed.

Table 4. Vegetation Canopy Ratings

Cover (%)	Cover rating
> 61	4
21-60	3
2-20	2
< 2	1
None	0

- Step 6. For those plant beds where invasive species are present, rake throw sampling should be conducted to quantify the degree of infestation. The number of rake throws required depends upon the homogeneity of the plant bed. In plant beds highly dominated by one species, as few as three throws may be sufficient if the results are the same each throw. Alternatively if each rake throw has plant species compositions that vary, then a larger number of rake throws will be required (e.g., 4-5 throws).
- Step 7. Record the number of rows with information (from the **Species information** area) in the **Total # of Species** box at the top of the data sheet.
- Step 8. Return to the approximate center of the plant bed and record GPS derived latitude and longitude coordinates and record the coordinates in the **Site coordinates** area of the data sheet. (A map grid may also be used to determine latitude and longitude coordinates if GPS is not available.) If appropriate, also record the GPS derived latitude and longitude coordinates for the location that defines the furthest lakeward extent of the plant bed. Note the approximate locations of both points with an "X" on the attached plant bed map.
- Step 9. Repeat steps 1 to 8 for each plant bed surveyed. Remember to start a new data sheet for each new bed encountered.

5. Post Survey Analysis

All waterbody summary information and GPS metadata is recorded on the **Waterbody cover sheet**. Datasheets are completed to the greatest extent possible following the survey. The map is completed with all relevant information and plant beds drawn. The surface area (acres) of each plant bed is determined and recorded in the **Bed Size** box on the data sheet. When gross historical changes in species composition, dominant species, and surface coverage are observed from year to year, notes are added to the **Comments** section.

6. Data & Equipment Management

All data sheets are identified with the sampling organization's name and crew leader and recorder names. Photocopies are made of all data and log sheets. The photocopied data sheets are mailed to the Department of Natural Resources Division of Fish & Wildlife. All originals are retained by the sampling organization.

Endangered, threatened or rare species are recorded on the data sheet and approximate locations noted on the map through the use of the **Reference ID** box (See Step 4). The presence of such species should also be recorded on the Indiana Special Plant Survey Form (See Appendix A) and sent to the IDNR Division of Nature Preserves.

Voucher specimens are collected and directed to the attention of Dr. Robin Scribailo at Purdue-North Central.

To avoid the spread of exotic species, survey crews should insure that all traces of aquatic vegetation are removed from boats, motors, and sampling gear before surveying other lakes/streams.

7. References Cited

IDNR. 2004. Procedure manual for Tier II aquatic vegetation surveying. Indiana Department of Natural Resources, Division of Fish and Wildlife, Indianapolis, Indiana. 10p.

Yin, Y., Winkelman, J.S., and H.A. Langrehr. 2000. Long Term Monitoring Program procedures: Aquatic vegetation monitoring. U.S. Geological Survey, Upper Midwest Environmental Sciences Center, La Crosse, Wisconsin, LTRMP 95-P002-7. 8pp. + Appendices A-C.

APPENDICES

Aquatic Vegetation Reconnaissance Sampling

Waterbody Cover Sheet

Surveying Organization:

Waterbody Name:

Lake ID:

County:

Date:

Habitat Stratum:

Ave. Lake

Depth (ft):

Lake Level:

GPS Metadata

Crew

Leader:

Datum:

Zone:

Accuracy:

Recorder:

Method:

Secchi Depth (ft):

Total # of Plant

Beds Surveyed:

Total # of

Species:

Littoral Zone Size (acres):

Measured

Estimated

Littoral Zone Max. Depth (ft):

Measured

Estimate (historical Secchi)

Estimated (current Secchi)

Notable Conditions:

Quad Code: _____

Indiana Special Plant Survey Form

Element Name: _____

Surveyor (s): _____ Date: _____ Time: _____ to _____

Location: _____ ¼ _____ ¼ _____ ¼ _____ Sec. _____ T _____ R _____ Quad name: _____

Repeat visit: Yes No Repeat visit needed: Yes No When: _____

EO boundaries mapped: Yes No County: _____

Area name (if applicable) _____

Biology

<u>Phenology</u>	<u>Approx # Indiv</u>	<u>Population Area</u>	<u>Age Class</u>
__ In leaf	__ 1-10	__ 1 yd ²	__ % Seedlings
__ In bud	__ 11-50	__ 1-5 yd ²	__ % Immature
__ In flower	__ 51-100	__ 5-10 yd ²	__ % 1 st year
__ In fruit	__ 101-1000	__ 10-100 yd ²	__ % Mature
__ Seed Dispersing	__ 1001-10,000	__ 100 yd ² -2 ac	__ % Senescent
__ Dormant	__ 10,001+	__ 2 ac +	

Comments on above: _____

Compared to your last visit to this site: Approx # Indiv Population Area Age Class

__ more __ more __ same

__ same __ same __ diff

__ less __ less

Reproduction Is reproduction occurring? __ Type: __ sexual, __ asexual, __ both

Show exact location and boundaries of taxon on map. (attach)

Population Distribution ___ solitary, ___ clumps or dense groups, ___ small patches or cushions
 ___ small colonies or large carpets, ___ large, almost pure population stands.

Vigor: 1) very feeble, 2) feeble, 3) normal, 4) exceptionally vigorous

Evidence of symbiotic or parasitic relationships:

Habitat

<u>Aspect</u>	<u>Slope</u>	<u>Light</u>	<u>Topographic Position</u>	<u>Moisture</u>
___N	___Flat	___Open	___Crest	___Inundated (Hydric)
___E	___0-10'	___Filtered	___Upper slope	___Saturated(Wet-mesic)
___S	___10-35'	___Shade	___Mid-Slope	___Moist (Mesic)
___W	___35' +		___Lower slope	___Dry (Xeric)
	___Vertical		___Bottom	

Elevation: _____ ft to _____ ft. Surface Relief: ___/: ___∪: ___∩: ___—: ___~~

Substrate/Soils: _____

Associated Natural Community/Plant Community: _____

List other members of this genus co-occurring at this site: _____

Characteristic associated species: _____

Estimated size of potential Habitat: (as in population area) Boundaries mapped: yes no

Ownership info: (if known) _____

NOTE: Collect specimen if a healthy, viable population exists. Collection # _____

Appendix B.

Explanations of Fields on the Aquatic Vegetation Waterbody Cover Sheet

Surveying Organization	Name of agency, corporation, group, individual, etc. that is collecting the data
Waterbody name	Common name of the lake or stream. Name should be consistent with the name found on most maps of the given waterbody (e.g. Lake Lemon, not Lemon Lake).
Lake ID	Unique State assigned alphanumeric code for the specific waterbody. Available through IDNR, Division of Fish & Wildlife.
County(s)	Name of the county(s) where sampling was conducted. When the waterbody or stream section traverses more than one county, list the primary county (county with the greatest acreage of water) first.
Date	The month (MM), day (DD), and year (YYYY) on which a site was sampled. Zeros (0) must be written in so that the date has eight digits.
Habitat stratum	Each stratum code defines a unique, major aquatic geomorphic feature in the state of Indiana. The habitat stratum of the site according to the above protocol is an important ecological consideration, as well as, valuable for the purposes of stratifying future sampling. The letter codes are listed in Table 1.
Average Depth	Average depth of the lake. Reference bathymetric maps, state personnel, historic studies etc.
Lake Level	Lake level at the time of sampling
Crew leader code	The full name or ID number that uniquely identifies the individual responsible for certifying that the samples and the data on the form were collected in compliance with current protocol and are, to the best of their knowledge, complete and free of errors. This identifying field underscores the importance of above method and is an important chain-of-custody procedure.
Recorder code	A name or number or initials that uniquely identifies the individual recording the data on the data sheets.
Datum	One or more constants used for calculating positions or elevations. These series of constants are commonly referred to as NAD'83, NAD'27, WGS'84, etc.
Zone	The number that identifies the correct grid from which the coordinates were taken. All of the State of Indiana falls into Zone 16.

Accuracy	The GPS measure of possible error related to the geometry of satellites. This number value is recorded when the Lat/Long coordinates are recorded. The method field indicates whether the scale is PDOP (Percent dilution of precision) or FOM (Figure of Merit).
Method	<p>A code that identifies the method used to locate the site and the type of accuracy measurement used by the equipment.</p> <p>B = Base Map D = GPS with differential corrections and PDOP G = GPS without differential corrections and PDOP F = GPS with differential corrections and FOM X = GPS without differential corrections and FOM O = other (explain)</p>
Secchi Depth	Secchi depth is taken and recorded (feet) at a mid plant bed site as soon as depth allows and distance from shore is deemed appropriate.
Total # of Plant Beds	Number of plant beds surveyed on the particular lake/stream as part of this sampling effort.
Total # of Species	The total number of <u>unique</u> records (rows) in SPECIES INFORMATION on the data sheets from <u>all beds</u> . This number represents the species diversity for the entire waterbody.
Littoral Zone Size	Size (acres) of the entire littoral zone may be measured through a variety of mapping techniques or estimated by the surveyors. The method is then noted.
Littoral Zone Max. Depth	Maximum littoral depth may be measured at a variety of locations in the field and averaged <u>or</u> estimated through the use of current or historical Secchi disk data. The extent of the littoral zone can be determined by multiplying the average or current Secchi depth by three. The method is then noted.
Notable Conditions	Comments that describe any unusual weather or water quality conditions that may interfere with accurate sampling such as rain, strong winds, algal blooms, etc.

Appendix B. Explanations of Fields on the Aquatic Vegetation
Plant Bed Data Sheet

Organization name	Name of agency, corporation, group, individual, etc. that is collecting the data
Date	The month (MM), day (DD), and year (YYYY) on which a site was sampled. Zeros (0) must be written in so that the date has eight digits.

SITE INFORMATION

Waterbody name	Common name of the lake or stream. Name should be consistent with the name found on most maps of the given waterbody (e.g. Lake Lemon, not Lemon Lake).
Waterbody ID	Unique State assigned alphanumeric code for the specific waterbody. Available through IDNR, Division of Fish & Wildlife.
Plant Bed ID	Two-digit number assigned to uniquely identify each bed/site. Accuracy of the Plant Bed ID is critical because it links field data to be collected with data already available in the database. A zero must be written before the number so the ID # is a two-digit number starting with "01".
Substrate	A qualitative code assigned to substrate type following tactile and visual examination of sediment at the sampling site. Substrate is rated on a scale of 1 to 6 according to Table 5.
Marl	A "1" identifies the presence of a marl (calcium carbonate) sediment. The default is a "0".
High Organic	A "1" identifies the presence of coarse organic material in the sediment. The default is a "0".
Total # of Species	The total number of detail records (rows) in SPECIES INFORMATION that contain data on this particular data sheet.
Cover	<p>S = Percent canopy abundance of all submersed 'topped-out' species combined for the bed using the ratings described in Table 4.</p> <p>N = Percent canopy abundance of all nonrooted floating-leaved species combined for the bed using the ratings described in Table 4.</p> <p>F = Percent canopy abundance of all rooted floating-leaved species combined for the bed using the ratings described in Table 4.</p> <p>E = Percent canopy abundance of all emergent species combined for the bed using the ratings described in Table 4.</p>

SITE COORDINATES (Recorded when the approximate center of the plant bed is determined and the furthest lakeward extent is known.)

Latitude The latitude coordinate for the site (either center or extent). The coordinate is recorded via a GPS unit after plant bed boundaries are estimated.

Longitude The longitude coordinate for the site (either center or extent). The coordinate is recorded via a GPS unit after plant bed boundaries are estimated.

SPECIES INFORMATION

Species code The alphanumeric six letter code for a species. Most of the species codes are available in Appendix C. If the genus of a plant is known and species unknown, then a new code is made up with the first four letters of the genus name and a '?' (question mark) inserted between the second and third letters. For examples, "PO?TA" for *Potamogeton* sp., and "MY?RI" for *Myriophyllum* sp. Using the species code of a suspected species is preferable, however, when based on the suggestion of the vegetation specialist. The confidence level of identification will be reflected in the **QE** code.

Abundance A number (1-4) that represents the percent abundance of a particular species in the community at the bed/site using the ratings described in Table 2.

QE A number (0–3) used to flag the taxonomic identification uncertainty (Table 3).

Voucher A code denoting whether a voucher specimen was taken of the species.
 0 = no voucher taken
 1 = voucher taken, and not sent out for identification
 2 = voucher taken, and sent out for identification

Reference ID A number or letter that denotes a specific location of a species of concern. The number or letter is referenced on an attached map showing the approximate location(s).

Comments A field for recording weather (e.g., overcast, rain, sunny) and any additional observations. Limit comments to 100 characters.

Reminder Information (Abbreviated glossary of codes used in the data fields.)

Appendix C. Species Codes

Species Code	Scientific Name	Common Name	Vegetation Type
ALGA	Any species of filamentous alga (incl. Spyrogyra, Cladophora, Hydrodictyon)	algae	NV
AZ?OL	<i>Azolla</i> sp.	a mosquito fern sp.	NV
AZCA	<i>Azolla caroliniana</i>	Carolina mosquito fern	NV
AZME	<i>Azolla mexicana</i>	Mexican mosquito fern	NV
CACA	<i>Cabomba caroliniana</i>	fanwort	
CEDE4	<i>Ceratophyllum demersum</i>	coontail	SB
CH?AR	<i>Chara</i> sp.	a chara sp.	SB
CHAS	<i>Chara aspera</i>		SB
CHBR	<i>Chara braunii</i>		SB
CHBR2	<i>Chara brittonii</i>		SB
CHCO	<i>Chara contraria</i>		SB
ELCA7	<i>Elodea canadensis</i>	Canadian waterweed	SB
ELNU2	<i>Elodea nuttalli</i>	western waterweed	SB
LEMN	Species within the Lemnaceae	duckweeds	NV
LEMI3	<i>Lemna minor</i>	small or common duckweed	NV
LETR	<i>Lemna trisulca</i>	star duckweed	NV
LUDE4	<i>Ludwigia decurrens</i>	primrose-willow	FL
LVWORT	<i>Riccia</i> sp., <i>Ricciocarpus</i> sp.	a liverwort species	NV
MYSI	<i>Myriophyllum sibiricum</i>	northern watermilfoil	SB
MYS2	<i>Myriophyllum spicatum</i>	Eurasian watermilfoil	SB
MY?RI	<i>Myriophyllum</i> , unidentified species	a watermilfoil sp.	SB
NAFL	<i>Najas flexilis</i>	slender naiad	SB
NAGR	<i>Najas gracillima</i>	slender waternymph	SB
NAGU	<i>Najas guadalupensis</i>	southern waternymph	SB
NAMI	<i>Najas minor</i>	brittle waternymph	SB
	<i>Potamogeton foliosus</i> , <i>P. pusillus</i> ,		

NLPW	or other unidentified narrow-leaved pondweeds	narrow-leaved pondweeds	SB
NELU	<i>Nelumbo lutea</i>	American lotus	FL
NI?TE	<i>Nitella</i> sp.	a nitella sp.	SB
NOAQVG		no aquatic vegetation in site	NV
NULU	<i>Nuphar variegatum</i> (formerly <i>N. luteum</i>)	yellow pond lily	FL
NYTU	<i>Nymphaea tuberosa</i>	white water lily	FL
POAL8	<i>Potamogeton alpinus</i>	red or alpine pondweed	SB
POCR3	<i>Potamogeton crispus</i>	curly-leaf pondweed	SB
POEP2	<i>Potamogeton epihydrus</i>	ribbon-leaf pondweed	SB
POFO3	<i>Potamogeton foliosus</i>	leafy pondweed	SB
POGR8	<i>Potamogeton gramineus</i>	variable pondweed	SB
POIL	<i>Potamogeton illinoensis</i>	Illinois pondweed	SB
PONO2	<i>Potamogeton nodosus</i> (formerly <i>P. americanus</i>)	American pondweed	SB
POPE6	<i>Potamogeton pectinatus</i>	sago pondweed	SB
POPR5	<i>Potamogeton praelongus</i>	white-stemmed pondweed	SB
POPU7	<i>Potamogeton pusillus</i>	small pondweed	SB
PORI2	<i>Potamogeton richardsonii</i>	Richardson's pondweed	SB
POZO	<i>Potamogeton zosteriformis</i>	flat-stemmed pondweed	SB
RAFL	<i>Ranunculus flabellaris</i>	yellow water-cup (yellow water buttercup)	SB
RALO2	<i>Ranunculus longirostris</i> (incl. <i>R. trichophylus</i>)	white water-cup (rigid white water buttercup)	SB
SACU	<i>Sagittaria cuneata</i>	Northern arrowhead	
SPPO	<i>Spirodela polyrhiza</i>	greater duckweed	NV
UNKN01		Unknown specimen No. 1	
UNKN02		Unknown specimen No. 2	
UTMA	<i>Utricularia vulgaris</i> (also known	common bladderwort	SB

	as <i>U. macrorhiza</i>)		
VAAM3	<i>Vallisneria americana</i>	wild celery	SB
WO?LF	<i>Wolffia</i> , unidentified sp.	a watermeal sp.	NV
WOCO	<i>Wolffia columbiana</i>	watermeal	NV
ZAPA	<i>Zannichellia palustris</i>	horned pondweed	SB
ZODU	<i>Zosterella dubia</i> (also known as <i>Heteranthera dubia</i>)	water stargrass	SB